

The Arima-HiC Kit and Service for Chromosome Spanning Assemblies

PRODUCT INFOSHEET

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Introduction

The availability of high-quality reference genomes has had a profound impact on the understanding of genome function and species evolution. Recent years have seen a rapid expansion of long-read and long-range methods, including Hi-C^{1,2}, a NGS-based assay that preserves chromosome-range information during sample preparation.

Hi-C sequencing data uses the preserved chromosome-range information to transform contigs to chromosomes. Despite its utility, broad adoption of Hi-C has been plagued by labor-intensive complex protocols, prolonged workflow durations, inconsistent experimental results, excessive sequencing requirements, and expensive Bioinformatics analyses/services.

The Arima-HiC Kit overcomes these technical and economical limitations with the development of a highly simplified and robust protocol that streamlines Hi-C to a 6-hour, 8-step procedure, followed by library prep and NGS. The Arima-HiC sequencing data is then used to scaffold contigs to generate chromosome-spanning assemblies via open-source software such as SALSA^{3,4} and 3D-DNA⁵.

These key advancements have persuaded researchers, including the Vertebrate Genomes Project (Fig.1), to use Arima-HiC to generate high-quality assemblies of hundreds of species across plant and animal kingdoms.

Highlights

Fast and User-Friendly Workflow

- Rapid time to Hi-C libraries with minimal hands-on touch-points, 6-hour prep time (1 hour hands-on time)
- Democratize analysis workflow by using well-documented open source tools such as SALSA (<https://github.com/machinegun/SALSA>) for chromosome-spanning assemblies

Proven Performance

- Obtain best-in-class inter-contig Hi-C signal across sample types
- Innovative multi-restriction enzyme chemistry improves overall genome coverage
- Generate uniform "spread" of inter-contig HiC signal to scaffold contigs regardless of gap sizes

Assured Quality

- Arima-HiC is validated in a wide-range of sample preservation and transportation conditions through its selection by the Genome10K consortium to generate high-quality assemblies of 260 species
- Assured library quality to ensure sequencing and assembly success with quantitative and predictive and QC steps
- Obtain both long-range sequence and chromatin conformation information with one assay

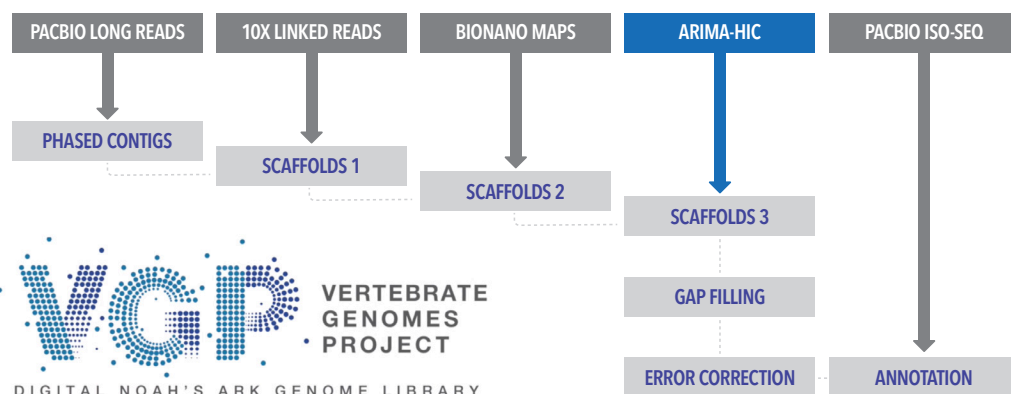


Figure 1: Arima Genomics is a technology partner of the VGP Phase 1. Long-read PacBio sequencing is performed to generate the initial contigs, followed by long-range scaffolding approaches to assemble contigs into chromosomes.

Image courtesy of Vertebrate Genomes Project

Fast and Easy Workflow

The Arima-HiC workflow was optimized to enable first time Hi-C users to generate high-quality data with ease (Fig.2). The rapid 6-hour protocol limits prolonged exposure of chromatin to external agents, leading to significant enrichment of inter-contig signal.

The use of a unique combination of multiple 4-base cutting restriction enzymes (RE) for chromatin digestion results in greater spread (per-base uniformity) of inter-contig signal to assemble all contigs regardless of gap-sizes.

Overall, the Arima-HiC Kit was designed to maximize the ease-of-use, with minimal total time and hands-on steps, compatibility with downstream library prep kits for Illumina NGS, 96-well plate compatible design, and support for a broad range of sample types and species (Table 1). Upon sequencing of the Arima-HiC libraries, open source Bioinformatics tools such as SALSA^{3,4} and 3D-DNA⁵ can be used to map Arima-HiC data to contigs to scaffold them into accurate chromosome-spanning assemblies.

In addition to scaffolding, Arima-HiC data has also been used to polish individual bases of the contigs as Arima-HiC is sequenced on an Illumina NGS³ (Table 1).

Proven Performance

Rigorous external and internal testing resulted in an Arima-HiC Kit with robust performance. When key opinion leaders compared Arima-HiC data with Hi-C data generated by competitors, the Arima-HiC data manifested higher inter-contig signal (Fig.3A). Importantly, the competitor's Hi-C data manifested a rapid drop in inter-contig signal with increase in contig gap-sizes. Arima-HiC data, on the other hand, manifested 2-3 fold higher inter-contig signal regardless of the contig gap-size (Fig.3B).

Subsequent SALSA^{3,4} analyses of Arima-HiC data converted 4Mb Oxford Nanopore contigs to 125Mb NG50 chromosome-spanning human genome assemblies, with as little as 30X Arima-HiC sequencing depth³ (Fig.4). This is an extraordinary performance given that even the

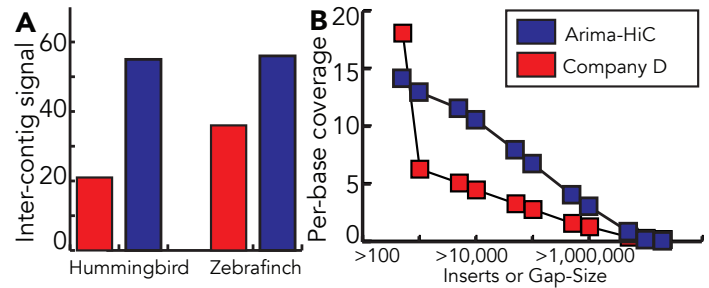


Figure 3: Arima-HiC data generates higher inter-contig signal strength and spread, critical features that enable chromosome-spanning assemblies at reduced sequencing.

(A) Arima-HiC data and Competitor's Hi-C (data generated by Company D) is mapped to Hummingbird and Zebrafinch contigs generated by Pacific Biosciences Sequencers. Regardless of the species, Arima-HiC consistently generates higher inter-contig signal. (B) Hummingbird Hi-C datasets from Arima Genomics and Competition analyzed in the context of insert-sizes. That is, when Hi-C reads are categorized by insert-sizes, Arima-HiC maintains high signal (2-3 fold) & coverage regardless of insert-size. In the context of assembly, this "spread" of signal can enable contigs of all gap-sizes to be well-assembled, to generate accurate assemblies at reduced sequencing cost. To enable an unbiased analyses, these were performed with 2M Hi-C reads. Analyses performed by Arang Rhie (NIH). Shared with permission.

"Arima was selected for Phase I (of the VGP) based on the quality of their data, proven by their ability to generate reproducible and high-quality data despite variability in input sample quality and quantity."

- Gene Myers PhD, Director of Max Planck-CBG and G10K Council Member

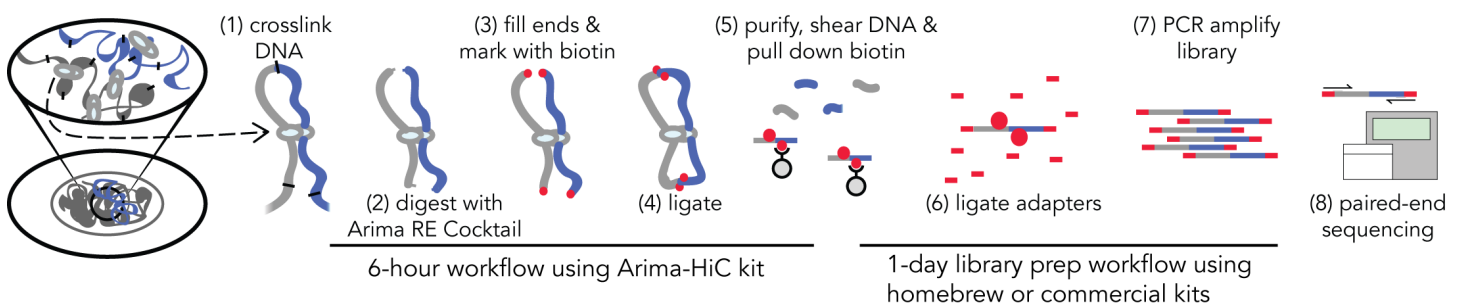


Figure 2: The Arima-HiC workflow results in ligated and biotinylated DNA that is PCR-amplified and prepared as a library using a multitude of library prep kits with appropriate adapters for paired-end Illumina sequencing.

GRCH38 reference assembly generated by cost-prohibitive combination of multiple technologies had an NG50 of only ~140Mb, suggesting that the high quality of Arima-HiC libraries and sequencing data offers substantial technical and economic benefits to the user. These benefits can be leveraged via Arima-HiC kits or services (sample to assembly).

Additionally, computational estimations suggest that about one-third of the genome (estimated from chr1 of the human genome) is inaccessible when Hi-C is performed with a 4-base restriction enzyme (RE), resulting in potentially limited interaction signal for one-third of genomes. Arima-HiC overcomes this limitation by using a unique RE cocktail and generates near complete genome accessibility.

Demonstrated Utility

The ease-of-use, consistency, and proven performances of Arima-HiC workflow, and, the open-source Bioinformatics strategy have made the Arima-HiC kits and services as a highly popular choice for generating chromosome-spanning assemblies. Indeed, Arima-HiC has been selected by the Genome10K consortium to generate high-quality assemblies and base-polishing of >260 vertebrate species⁶.

An additional benefit is that the same Arima-HiC data generated for genome scaffolding can then be used to investigate the 3D conformation of the genome. Chromatin conformation information is useful for many studies including cell development, gene regulation, and pathogen resistance.

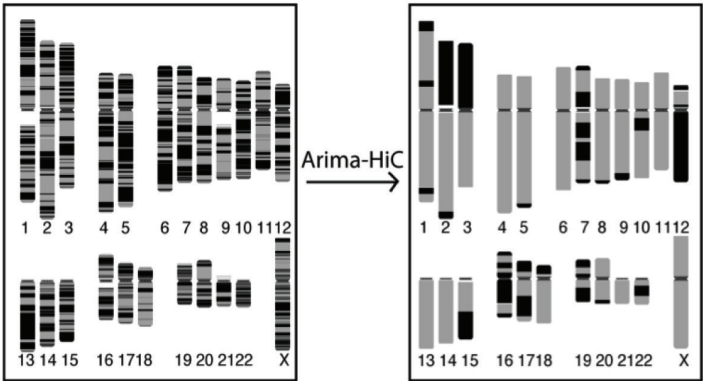


Figure 4: 125Mb NG50 Human chromosome-spanning assemblies, generated by Arima-HiC data 4Mb contigs from Oxford Nanopore converted to 125Mb assemblies via Arima-HiC data, using SALSA open-source tool. Change in color in the ideogram represents a contig gap or error. Post Arima-HiC assemblies have minimal color change, suggesting long (NG50 125Mb and chromosome-spanning) & accurate assemblies. Figure from Ref (3), shared with permission.

Additional Details

Please refer to the Genome Conformation Application Note, available by contacting info@arimagenomix.com.
Learn more online at arimagenomix.com

References

1. Lieberman-Aiden E, et al "Comprehensive Mapping of Long-Range Interactions Reveals Folding Principles of the Human Genome" Science 326, 289-293 (2009)
2. Rao SP, et al "A 3D Map of the Human Genome at Kilobase Resolution Reveals Principles of Chromatin Looping" Cell 159, 1665-1680 (2014)
3. Ghurye J, et al "Integrating Hi-C links with assembly graphs for chromosome-scale assembly" BioRxiv, doi: <http://dx.doi.org/10.1101/261149> (2018)
4. SALSA Bioinformatics Tool for Arima-HiC. <https://github.com/machinegun/SALSA>
5. Dudchenko O, et al "De novo assembly of the Aedes aegypti genome using Hi-C yields chromosome-length scaffolds" Science 356, 92-95 (2017)
6. Genome10K Press Release (1). <https://www.businesswire.com/news/home/20180117006561/en>

Table 1: Arima-HiC Specifications

Total Time	6 hours
Hands-on Time	1 hour
Number of Steps	8
Automation Capability	Single-tube, 96-well plate compatible
Restriction Enzymes (RE)	RE cutting at GATC and GATC
Per-base Genome Uniformity "Spread" (fraction of genome with avg. seq. depth)	~90%, as estimated from Human genome. Similar performance expected in all species.
Sample Types	Seeds, Tissue, blood, cell lines, whole insects
Sample Storage Conditions	Fresh/frozen, Cross-linked, Ethanol
Input Quantity	Amount of sample manifesting ~1ug DNA, low input protocols also available
Species	Plants, Invertebrates, Vertebrates
Library Prep Compatibility	KAPA Hyperprep, Swift Accel NGS 2S, Illumina TruSeq, NEBNext Ultra II, others
NGS Compatibility	Illumina NGS paired end reads
Library Complexity	1 reaction provides libraries complex enough for 600M reads
Sequencing Depth (X), recommended	Arima-HiC: 15-60X, depending on the contig NG50
Data Analysis	SALSA tool (Ref 3) and other open source tools recommended



Dwarf Crocodile



Salamander



European Turtle Dove



Hummingbird



Common Yellowthroat



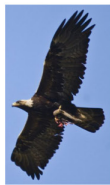
Italian Sparrow



Long-tailed Finch



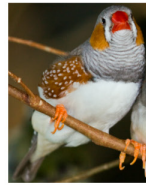
Kakapo



Golden Eagle



Blue-capped Cordonbleu



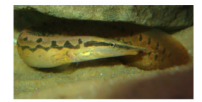
Zebra Finch



Chicken



Reedfish



Tire Track Eel



Greenland Shark



Mexican Tetra
(Blind Cave Fish)



Japanese Pufferfish



Indian Glassy Fish



Blunt-snouted Clingfish



Climbing Perch



Atlantic Cod



Gilt-head Bream



Burbot



Zebrafish



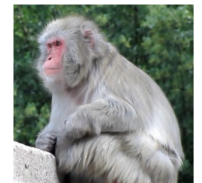
Turquoise Killifish



Eurasian Red Squirrel



Pale Spear-Nosed Bat



Macaque



Canadian Lynx



Cow



Horse



Mouse



Horseshoe Crab



Ant



Mosquito



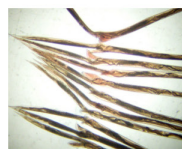
Bumblebee



Drosophila melanogaster



Schmidtea mediterranea



Barber's Pole Worm



Starlet Sea Anemone



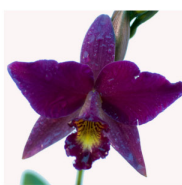
Wheat



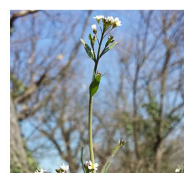
Brachypodium



Tobacco



Orchid



Arabidopsis

A Representation of the Plant and Animal Species Being Analyzed via the Arima-HiC Workflow



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